

Dual Effect of Alternariol on Acetyl Cholinesterase and Monoamineoxidase Extracted from Different Parts of Rat Brain

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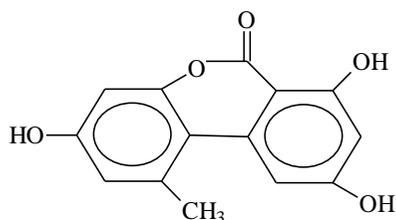
ABSTRACT Alternariol is a metabolite product of different strains of *alternaria tenuis* fungus. Its structure has a similarity to cannabinol derivatives and it has a chemical formula 3,4,5 trihydroxy 6-methyl dibenzoxy- α -pyrone. The effect of the alternariol on both acetylcholinesterase (AChE) and monoamineoxidase (MAO-A) enzymes extracted from whole and five different parts of male albino rat brains; namely: frontal cortex, basal ganglia, cerebellum, pons, medulla oblongata was studied. Kinetic studies were done to determine the type of inhibition of AChE and MAO-A enzymes and the enzyme –inhibitor dissociation constants (Ki) by alternariol. The results indicated that alternariol inhibited both AChE and MAO –A enzymes of the cortex and medulla oblongata more than the extracts of the other parts of the brain. These parts are responsible for perception, motor, sensory, psychic activities and reflex centers of respiration. The inhibition of these enzymes increased with increasing the amount of alternariol added to the assay mixture, i.e., the inhibition was dose dependent and of the competitive type. The values of Ki for alternariol – AChE enzyme extracts varied from 12.0 to 15.0 mmol/L and for alternariol – MAO enzyme extracts varied from 28 to 30 mmol/L and were of the same order of magnitude. The difference in the degree of inhibition of the extracts of these brain parts could be attributed to the slight difference in the structure i.e; arrangement of their amino acids (isozyme phenomenon) and to their distinct gene loci. The inhibition of AChE and MAO-A by alternariol may save ACh and biogenic amines which are of great importance for the patients suffering from Alzheimers and dementia.

Keywords: Acetylcholine; 5 Hydroxytryptamine (Serotonin); Alternariol; Acetyl Cholinesterase; Monoamine Oxidase.

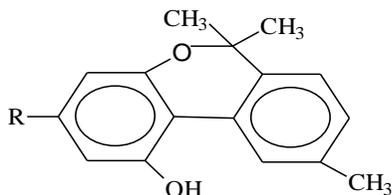
INTRODUCTION

Alternariol is a metabolite produced by trihydroxy 6-methyl dibenzoxy - α -pyrone different strains of *alternaria tenuis*. [1] Its $C_{14}H_{10}O_5$ [I]. It has striking similarity to structure was established as 3,4,5 cannabinal derivatives [II].

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I



II

Several reports [2,3] have attempted to correlate the behavioral effects of tetrahydrocannabinol (Delta 8 THC) with changes in the level of 5-hydroxytryptamine (serotonin) or its metabolite. THC is an important psychoactive ingredient in marijuana [4], which is the most widely illegal recreational drug in USA.

Our preliminary experiments showed that alternariol manifested inhibitory effect on whole rat brain acetylcholinesterase (AChE) and monoamine oxidase (MAO) activities [5].

Cholinesterases are a group of enzymes that degraded the esters of choline and play a role in neurotransmission in the autonomic and somatic motor nervous system. Their inhibition results in accumulation of acetylcholine. AChE is used as a human biomarker of organophosphorus pesticides

exposure [6].

Monoamine oxidase E.C. 1.4.3.4 (MAO) is an enzyme that oxidizes mono amine neurotransmitters and neuro-mediators as well as exogenous bioactive amines [7,8].

Inhibitors of MAO-A have shown to be effective antidepressants, and have been shown to be of value in the treatment of Parkinson's disease [9].

The present study was conducted to investigate the inhibitory effect of alternariol on AChE and MAO-A isozymes, extracted from whole and five parts of rat brain, namely; frontal cortex, basal ganglia, cerebellum, pons and medulla oblongata.

Enzyme kinetic studies were done to determine the type of inhibition and enzyme inhibitor dissociation constant (K_i) of AChE and MAO-A by alternariol, and to know

which of these parts was inhibited by alternariol more than other parts.

MATERIAL AND METHODS

Alternariol was prepared by growing a strain of *alternaria tenuis*, Catalogue number S.M. 108, on Czepek-Dox medium [1] using either glucose or molasses, as a carbon source [1]. The metabolite was extracted from the dried defatted mycellian with ether, then purified by repeated crystallisation from dioxane giving colorless needles m.p. 350° (decomp).

Chemicals: were purchased as follows: acetylthiocholine iodide (AThChI) from BDH chemical,Ltd, Poole (England) Dithio-bis-nitro benzoic acid from Aldrich chemical Co.Ltd Gillingham, England. 5-hydroxytryptamine creatinine sulphate (contains 43.5% of 5HT, serotonin) from May and Baker LH, Dagenham (England)

Animals: Fifty male albino rats (weighing 100-150 g) aged 2 months were used in the experiments .Rats were supplied from the Medical Research Institute animal house, Alexandria University, Egypt. Rats were housed in group cages (5 in each cage) and

allowed free access of food and tap water. The rats were killed by decapitation after subjected to an overnight fast, with free access of water. Each brain weighed about 1.6 g.

Preparation of AChE from whole and different parts of rat brain:

Whole brain as well as the different parts obtained from 10 g brain tissues were isolated, weighed and homogenized in ice cold phosphate buffer (0.1 mol / L, pH 8.0) [10]. The resulting homogenates were centrifuged twice at 600×g for 10 min to remove cellular debris. The supernatant was centrifuged at 10,000 ×g for 20 min, and the resulting precipitate was suspended with the same buffer and used as a source of the enzyme in the assay.

AChE assay: The activity of AChE was measured by the method of Ellman et al [11].The assay mixture contained: 2.0 ml phosphate buffer (0.1 M , pH 8.0) containing an amount of brain tissue equivalent to 0.1 unit of AChE, 100 ul dithio bis nitro benzoic acid (DTNB,0.1 mol/L) and 20 ul acetyl thiocholine iodide (AThChI, 21.67 mg/ml).

The type of inhibition and the enzyme – inhibitor dissociation constant (K_i) of AChE were measured at different concentrations of alternariol: 0 (control), 5.0, 10.0 and 15.0 mmol/L, while AThChI concentrations were varied, (2.5, 5.0, 7.5 and 10.0 mmol/L) for each concentration of alternariol. AThChI and alternariol were added simultaneously to the assay mixture.

Preparation of MAO-A from whole and different parts of rat brain: The MAO enzymes were prepared from whole and different parts of rat brain as described before [12,13,14]. Ten brains out of 40 were used (15g) as whole brain enzyme preparation. From the 30 other brains, the appropriate parts were isolated, washed with ice cold saline and weighed each separately. These parts were the frontal cortex (15g), basal ganglia (8g), pons (7g), medulla oblongata (10g) and cerebellum (15g). Whole brain as well as the different parts of the brain were homogenized in ice cold phosphate buffer Na_2HPO_4 : NaH_2PO_4 , PH 7.4, 0.1 mol/L containing 0.25 mol/L sucrose. The

homogenates were centrifuged at 12000xg for 20 minutes. The resulting supernatants from the second centrifugation were discarded, and the mitochondrial precipitates were resuspended in the same phosphate buffer to give a final protein concentration of 10 mg/ml. The protein content was determined by Lowry's method [15].

MAO assay The activity of MAO-A was measured by the method of Udenfreind et al [16,13,14]. The assay mixture contained: 1ml 5HT (166 $\mu\text{mol/ml}$) and 1ml enzyme containing 10 mg protein in phosphate buffer. After incubation at 37°C for 1hr, 1ml of each of 1-nitroso, 2-naphthol (0.1% in 95% ethyl alcohol) and acid nitrite (freshly prepared), reagents were added to react with the unhydrolyzed 5HT. Measurements were carried out at 540 nm. The type of inhibition and K_i of MAO-A were measured at different concentrations of alternariol: 0 (control), 5, 10 and 15 mmol/L while 5HT concentrations were varied. (111, 166, 222 and 333 $\mu\text{mol/L}$) for each concentration of alternariol. 5HT and alternariol were added simultaneously to the assay

mixture.

RESULTS

The activity of AChE in whole and different parts of rat brain was measured at constant substrate concentration (0.5 mmol/L AThChI). It has been found that the basal ganglia extract possessed the highest specific activity. The values of K_m obtained from Lineweaver-Burk plot [17] of $1/V$ versus $1/S$ in the case of AChE extracts varied between 0.12 and 0.17. (table 1)

The activity of MAO in whole and different parts of rat brain was measured at constant substrate concentration (166 μ mol/L 5HT). It has been found that the basal ganglia and frontal cortex extracts possessed the highest activity of MAO. The values of K_m in case of MAO-A extracts varied between 0.28 and 0.33 mmol/L (table 2).

A double reciprocal plot of velocity versus substrate concentrations in the absence and presence of alternariol gave curves of the

competitive type for the inhibition of both AChE and MAO (fig. 1 and 2). This was confirmed by Cleland replot [18] of the slopes of the lines of (fig 1 and 2) versus inhibitor concentrations taking basal ganglia for AChE and MAO as examples (fig. 1 and 2).

The value of enzyme-inhibitor dissociation constant (K_i) of alternariol with whole brain AChE extract was 1.8mmol/L, and with different parts of the brain varied from 1.6 to 2.1mmol/L.

The values of K_i of alternariol with whole brain MAO-A extract was 2.4mmol/L and with the extracts of different parts varied from 2.4 to 3.8mmol/L.

The results also indicated that the highest affinity (K_i/K_m) of alternariol was with the extracts of AChE obtained from frontal cortex and medulla oblongata (13.12 and 13.33mmol/L) and to MAO-A extracts of frontal cortex and medulla oblongata also (11.88 and 11.20)

Table 1: Rate of hydrolysis of acetyl thiocholine iodide by acetyl cholinesterase extracted from whole and different parts of rat brain under the effect of alternariol. Values represent mean \pm S.D. of 3 repeated experiments.

Part of the brain	AChE activity umol/min/g wet wt	(a) Km m mol/L	(b) Ki m mol/L	(c) Ki/Km
Whole brain	18.4 \pm 0.12	0.15	1.8	12.0
Frontal cortex	15.2 \pm 0.09	0.16	2.1	13.12
Basal ganglia	27.8 \pm 0.18	0.14	1.8	12.85
Pons	14.3 \pm 0.10	0.15	1.7	11.33
Medulla oblongata	16.5 \pm 0.12	0.12	1.6	13.33
Cerebellum	5.5 \pm 0.07	0.17	2.0	11.76

a) Km, Michaelis constant.

b) Ki, enzyme inhibitor (alternariol) dissociation constant.

c) Ki/Km:affinity constant

Table 2: Rate of hydrolysis of 5-hydroxy tryptamine by monoamine oxidase extracted from whole and different parts of rat brain under the effect of alternariol. Values represent mean \pm SD of 3 repeated experiments.

Part of the brain	MAO activity umol/L 5HT hydrolyzed/mg protein/hr	(a) Km mmol/L	(b) Ki m mol/L	(c) Ki/Km
Whole brain	135 \pm 6.7	0.30	2.4	8.0
Frontal cortex	138 \pm 5.3	0.32	3.8	11.88
Basal ganglia	142 \pm 7.8	0.33	3.2	9.69
Pons	132 \pm 6.9	0.30	3.0	10.0
Medulla oblongata	104 \pm 5.8	0.25	2.8	11.2
Cerebellum	124 \pm 4.4	0.30	3.0	10.0

a) Km, Michaelis constant.

b) Ki, enzyme inhibitor (alternariol) dissociation constant.

c) Ki/Km:affinity constant

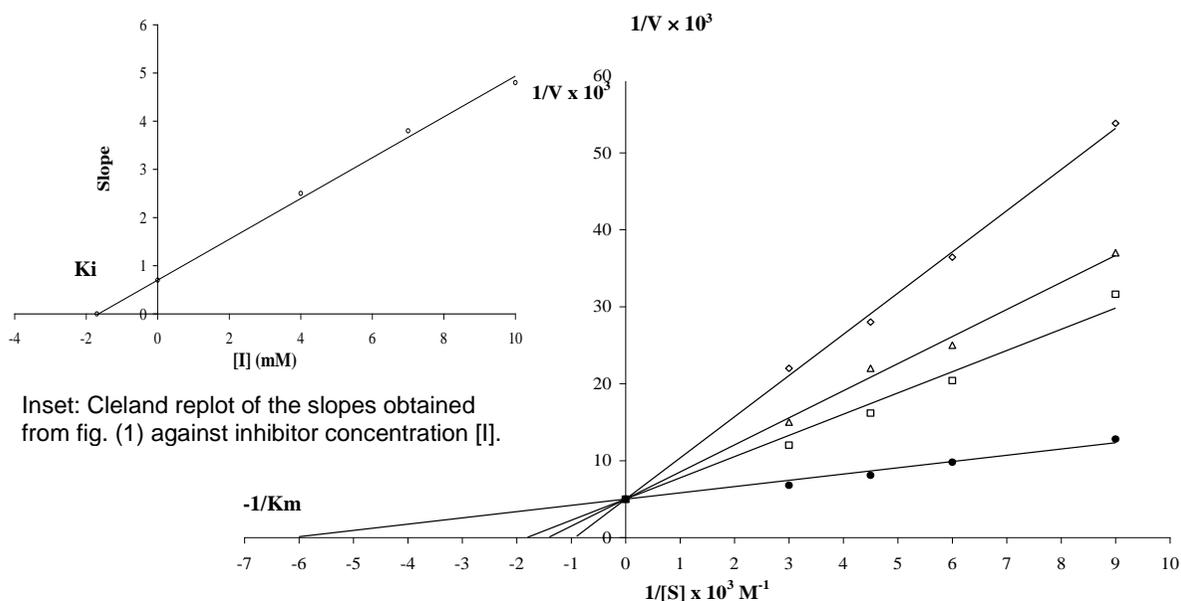


Figure (1): Competitive inhibition, obtained from Lineweaver-Burk plot of $1/V$ versus $1/S$, under the effect of alternariol on the rate of hydrolysis of AThChI by frontal cortex (AChE) extract. (●) Control, (◻) 5.0, (△) 10.0 & (○) 15mmol/L

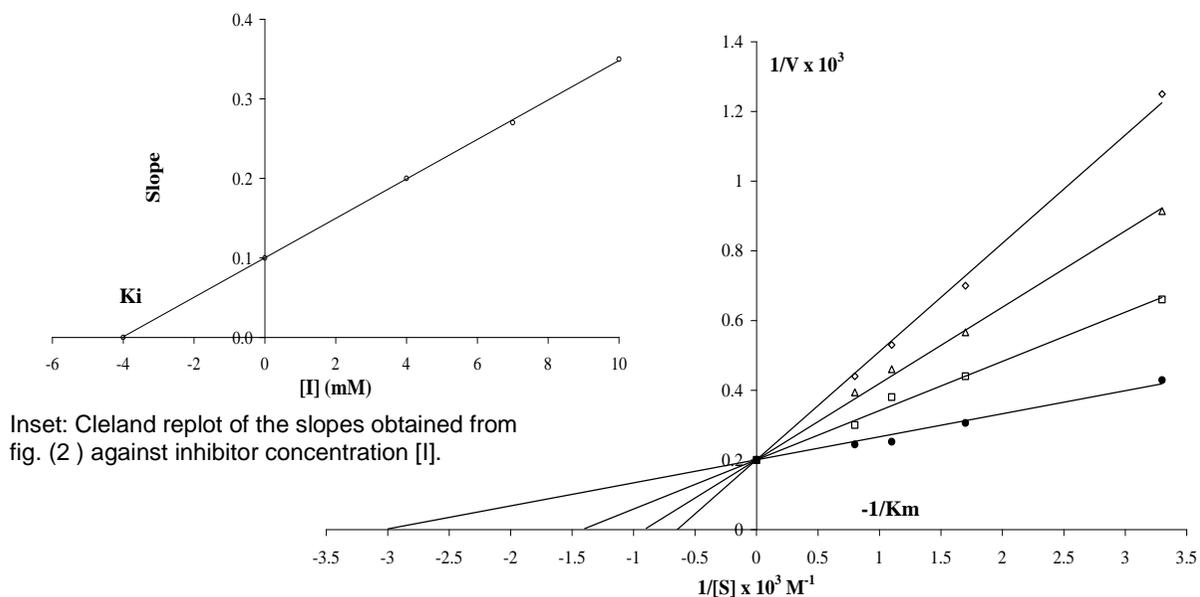


Figure (2): Competitive inhibition obtained from Lineweaver-Burk plot of $1/V$ versus $1/S$, under the effect of alternariol on the rate of deamination of 5 HT by frontal cortex (MAO) extract (●) Control, (◻) 5.0, (△) 10.0 & (○) 15mmol/L.

Discussion

The differences in the activity and enzyme-substrate dissociation constants (K_m) values, between the extracts of the parts of the brain in case of both AChE and MAO-A extracts are attributed to isozyme phenomenon [19].

A-Effect of Alternariol on brain AChE:

The value of enzyme-inhibitor dissociation constant (K_i) of alternariol with whole brain AChE extract was 1.8mmol/L, and with different parts of the brain varied from 1.6 to 2.1mmol/L. When comparing the value of K_i obtained in the reaction of alternariol with brain AChE with those previously obtained in the reactions of some biogenic amines, namely; 5-HT (18mmol/L), epinephrine (E) and norepinephrine (NE) (12.2 and 8.5mmol/L), the present results indicate that alternariol is 10, 7.0 and 5.0 times more potent than 5-HT, E and NE- respectively -in its inhibitory effect on AChE. [20,21]

Alternariol possessed higher affinity and binding to the enzyme extracts of

medulla oblongata than with the extracts of the other parts. This means that it affects the areas of perception, psychic activities and the reflex centers for respiration.

Alternariol is more effective on AChE than cannabinoids (Δ -8-THC and 11-hydroxy Δ -9-THC). These compounds showed only slight changes on AChE isolated from synaptic membranes [4]. In connection with the present results, Eubanks et al [22] demonstrated that Δ -9-THC, the active component of marijuana (which has structural similarity to alternariol) competitively inhibited AChE activity.

Effect of Alternariol on brain MAO-A

MAO-A inhibitors have shown to be effective antidepressants and of value in the treatment of Parkinson's disease [9]. As MAO-A metabolizes 5-hydroxytryptamine (serotonin) and noradrenaline (nor epinephrine), and is a neural enzyme, it was only natural that most drug research emphasized on MAO-A inhibitors.

In the present work, the lowest K_m values of MAO-A using 5HT as substrate was obtained with the extract of medulla oblongata 0.25mmol/L, showing that the substrate has more affinity to the enzyme extracted from M.O than to the extracts of other parts.

The lowest K_i value was obtained in case of the reaction of alternariol with MAO-A extract of medulla oblongata (2.8mmol/L) indicating that alternariol has more affinity to the active site of this extract than to the extracts of the other parts. When comparing the value of K_i obtained in the reaction of alternariol with MAO-A (2.4 mmol/L) with that previously obtained [14] with the reaction of acetylcholine with MAO-A (3.7 mmol/L), the results indicate that alternariol is 1.5 times more potent than acetylcholine in its inhibitory effect on MAO-A.

Previous reports (2) mentioned that delta- 9 THC (which is one of the main psycho-active components of cannabis)

has drastic effect on isolated rat liver mitochondria, detectable in the form of damage to cristae and outer membranes as well as changes in the respiration and ATPase activity. These findings coincide with the action of alternariol on MAO-A which is a mitochondrial enzyme (similar to ATPase).

The results showed that the degree of its inhibitory effect was varied between the extracts of the different parts of the brain. The lower K_i values of alternariol with MAO-A is markedly with the extract of medulla oblongata which is responsible for respiration. It is note-worthy to mention that the values of K_i obtained in cases of alternariol-AChE reactions were about half the values obtained in case of the reaction of alternariol with MAO-A extract. This indicates that the affinity of alternariol to AChE enzyme extracts was higher than its affinity towards MAO-A enzyme extracts. Moreover, both AChE and MAO extracts of the medulla oblongata were inhibited by

alternariol than the extracts of other brain parts. However, the difference in the values of K_m and K_i suggesting the existences of different substrate binding site of both enzymes: AChE and MAO [14].

All these discrepancies indicate that one amino acid can be responsible for binding of some (but not all) substrates and inhibitors [14,23], or due to the sequences of amino acids in the enzyme. The dual inhibition of AChE and MAO by alternariol may save AChE and the biogenic amines which are of great importance for the patients suffering from Alzheimers and dementia.

REFERENCES

1. Raistrick H, Stickings C.E, Thomas, R. Biochemistry of microorganisms: Alternariol and alternariol monomethyl ether, metabolic products of *alternaria tenuis*. *Biochem J* 1953; 55: 421-33.
2. Pal B, Ghosh JJ. Δ -1 tetrahydrocannabinol effects on the urinary excretion of 5-hydroxyindol acetic acid. *Biochem. Pharmacol.* 1972; 21:263-5
3. Bornheim LM, Lasker JM and Rawc JL. Human hepatic microsomal metabolism of delta 1-tetrahydrocannabinol. *Drug metab. Dispos* 1992; 2: 241-6.
4. Moore C, Rana S, Coulter C. Simultaneous identification of 2-carboxy-tetrahydrocannabinol, tetrahydrocannabinol and cannabiniol in oral fluid. *J Chromatogr B Analyt Technol Biomed Life Sci* 2007;852(1-2):459-64
5. Osman H M.Y., Osman M.Y. Dual effect of alternariol on acetylcholinesterase and monoamine oxidase extracted from different parts of rat brain. *FASEB* (2008), Abstract#564.
6. Nigg HN, Knaak JB. Blood cholinesterase as human biomarkers of organophosphorus pesticide exposure. *Rev Environ Contam Toxicol* 2000;136:11-29.
7. Youdin MBH and Fingberg JPM. New directions in monoamine oxidase A and B selective inhibitors and substrates. *Biochem. Pharmacol* 1991; 41: 155-162.
8. Yu PH. In: *neuro methods v; Neurotransmitter Enzymes* (Eds. Boulton A.A. Baker G.B and Yu PH). Humana Press. Clifton. NJ 1986; PP, 235-272
9. Tipton KF. *Biochemical and Pharmacological Aspects of Depression* (Eds. Tipton KF and Youdin MBH). Taylor and Francis, London 1989 pp.1-24
10. Osman MY, Abdel Tawab SM, Sharaf IA. Effect of physostigmine on cholinesterase activity in different parts of rat brain. *Arzneim. Forsch/Drugs Res.* 1995; 6: 663-5.
11. Ellman GI, Courtney KD, Andress, V, Featherstone RM. A new and rapid colorimetric determination of acetylcholinesterase activity. *Biochem. Pharmacol* 1961; 7, 88-95.
12. Roth J and Gillis C. Determination of β -phenylethylamine by monoamine oxidase. Inhibition by imipramine. *Biochem pharmacol* 1974;23:2537-40.

13. Mohamed YS, Osman MY, Mahfouz MM. Effect of acetylcholine on monoamine oxidase. *Biochem Pharmacol* 1976;22: 1665-7.
14. Osman MY, Osman HMY. The inhibitory effect of acetylcholine on monoamine oxidase A and B activity in different parts of rat brain. *Arzneim-Forsch* 2008, 58: 493-6.
15. Lowry OH, Rosebrough NG, Farr AL, Randall RJ. Protein measurement with the phenol reagent. *J Biol Chem.* 1951;193;265-9.
16. Udenfreind S, Weissbach H, Brodie BB. In *Methods of biochemical analysis* (ed. David Glick). Assay of serotonin and related metabolic enzymes and drugs. Vol. 6 Interscience London.1958;6:96-9.
17. Dixon M, Webb EC. *Enzymes*. 2nd ed. London: Longmans;1964:162. Cleland WW. *The enzymes* 3rd ed New York. Academic press: BoyerP, 1976; vol 2:1-33
18. Strench J. *Foundation of biochemistry*. New York and London, Plenum press, 1998 P 104.
19. Mohamed YS, Osman MY, Gabr Y. Inhibition of cholinesterase by 5-hydroxytryptamine. *Arzneim. Forsch (Drug Res)* 1975; 25:1.
20. Mohamed YS, Osman MY. Inhibition of cholinesterase by epinephrine and norepinephrine. *Arzneim Forsch (Drug Res)* 1978; 28 (1): 16-18.
21. Eubanks LM, Rogers CJ, Beuscher AE, Koob GF, Olson AI, Dikerson TJ, Janda KD. A molecular link between the active component of marijuana and Alzheimer's disease pathology. *Mol pharm* 2006; 3(6): 773-7.
22. Veselovsky HV, Vanov AS and Medvedev F. Is one aminoacid responsible for substrate specificity of monoamine oxidase A and B. *Vober Med Khim* 1998; 43:527-36.